pISSN 2508-1357, eISSN 2508-139X J Biomed Transl Res 2020;21(4):184-189 https://doi.org/10.12729/jbtr.2020.21.4.184 Received 17 Nov. 2020, Accepted 30 Nov. 2020

Original Article

Effects of microcystin on the motility of planarians

Young-Joo Yi1^{+*}, Yeon-Hwa Kim2⁺, A. A. Dilki Indrachapa Adikari¹, Seung-Tae Moon¹, Sang-Myeong Lee^{3*}

¹Department of Agricultural Education, College of Education, Sunchon National University, Suncheon 57922, Korea ²Division of Biotechnology, College of Environmental and Bioresource, Jeonbuk National University, Iksan 54596, Korea ³Laboratory of Veterinary Virology, College of Veterinary Medicine, Chungbuk National University, Cheongju 28644, Korea

Cyanobacteria (blue green algae) blooms formed in natural water resources due to the environmental pollution produce toxic compounds as secondary metabolites, causing health hazards to both humans and other living beings. Microcystin is a well-known toxin produced by cynobacteria. The present study was undertaken to evaluate varying concentrations and exposure times of two different forms of microcvstin, viz., -LR (MCLR) and -LA (MCLA), on the motility and seizure-like behavior of planarian (Dugesia japonica). Compared to control, reduced motility was observed in both the MCLR or MCLA treated groups, but did not differ significantly with increasing concentrations of microcystin. However, the number of seizure-like behaviors were increased dose-dependently in planarian exposed to MLCR or MCLA. Exposure time to microcystine also affected the motility and seizure-like behaviors of planarians; 24 hrs incubation with MCLR, and 48 and 96 hrs exposure to MCLA, showed significantly (p < 0.05) lower motility, as compared to the control. Assessing regeneration of the planarians revealed the simultaneous completion of eye formation at day 9 in planarians incubated in the absence or presence of MCLR or MCLA, thereby indicating that exposure to microcystin has no effect on the process. In conclusion, we determined that exposure to microcystins resulted in decrease in the number of motility, and induced abnormal behavior pattern in planarians. Further studies are required to identify the toxicity of microcystin that affects aquatic ecosystems.

Key words: cyanobacteria, microcystin, toxin, planarian,

motility

Introduction

Cyanobacteria, also known as the blue-green algae, have been present on earth for billions of years. They have continued to evolve and adapt to the changing environmental conditions in the modern world. These cyanobacteria are responsible for the oxygenic life of the earth due to their capability of photosynthesis [1]. They are known to thrive in both marine and fresh water sources. Evolution of the world has resulted in a negative adverse effect on the natural ecological systems. Anthropogenic activities, such as industrial revolution, improper agricultural activities, indiscriminate disposal of pollutants, and the construction of reservoirs [2], trigger proliferation of these bacteria. Moreover, global warming as well as the higher nutrient content and lower flow rate of the water resource, favors growth of these bacteria. Thus, blooming of these bacteria causes a striking greenish color in the water resource. It is very common to see blooming in tropical countries, since the environmental conditions are very conducive for cyanobacterial growth in temperate zones. Blooms of cyanobacteria are most prominent during late summer and early autumn, and may last for 2-4 months. This blooming adversely affects other living beings of the water resource such as aquatic fauna and flora, as well as humans using this water for their daily requirements [3-5].

Cyanobacteria belong to the gram-negative bacterial group, producing a vast range of toxic compounds as

[†]These authors contributed equally to this work.

^{*}Co-corresponding author: Young-Joo Yi

Department of Agricultural Education, College of Education, Sunchon National University, Suncheon 57922, Korea Tel: +82-61-750-3352, E-mail: yiyj@scnu.ac.kr

Sang-Myeong Lee

Laboratory of Veterinary Virology, College of Veterinary Medicine, Chungbuk National University, Cheongju 28644, Korea Tel: +82-43-261-3356, E-mail: smlee@chungbuk.ac.kr

secondary metabolites, which are mostly peptides (such as anabaenopeptins, aeruginosins, microcystins, nodularins, microginins or microviridins) and alkaloids [6]. Cyanobacteria toxin poisoning is widely reported in animals such as fish, swine, cattle, dog and bat [7]. Among the toxins produced by cyanobacteria (blue-green algae), microcystin has gained considerable attention due the ability of this substance to cause hepatotoxicity. Microcystin is a group of cyclic heptapeptide hepatotoxics that contain D- & L-amino acids, and hydrophobic amino acids [8]. The chemical structure consists of D-alanine, X, D-erythromethylaspartic acid (MeAsp), Z, 3-amino-9methoxy-2, 6, 8,-trimethyl-10-phenyldeca-4,6 dienoic acid (Ada), D-glutamic acid, methyldehydroalanine (Mdha) [9]. X and Z indicate variable amino acids and distinguish the molecules, for instance, microcystin-LR stands for leucine (L) and arginine (R) as respective X and Z amino acid. Currently, more than 90 variants with different toxicity of microcystins have been reported. Microcystin-LR is the most well known variant, and other common variants are microcystin-LA (X: leucine, Z: alanine), microcystin-RR (X: arginine, Z: arginine), microcystin-YR (X: tyrosine, Z: arginine) [10]. Symptoms of microcystin poisoning include diarrhea, vomiting, and facial paleness; in particular, it has been reported to damage the liver cell skeleton, causing cell necrosis and bleeding [11, 12].

The evaluation of microcystin toxicity is required to predict its effect on living organisms. Planarian is an invertebrate, flat animal that is easy to breed and manage, has both asexual and sexual reproduction, and a welldeveloped central nervous system similar to humans; hence, it has high value as an experimental animal and is excellent for application [13, 14]. This study was therefore conducted to investigate the effect of microcystin on the motility of planarians, as well as to determine whether it is possible as an alternative experimental animal model for environmental toxin evaluation.

Materials and Methods

Laboratory animals and reagents

The planarians used in this experiment (*Dugesia japo-nica*) were propagated and maintained in water at 18° C. Planarians were fed chunks of beef liver twice per week, and water was replaced with fresh drinking water after 1 hr feeding. At least one week prior to the experiment, feeding was discontinued. Microcystin-LR (MCLR) and Microcystin-LA (MCLA) used in the experiments were purchased from Sigma-Aldrich (#33893, Seoul, Korea) and Enzo (#ALX-350-096, Madison, NY, USA), respectively (Fig. 1). Each stock solution (100 μ g/mL) was prepared using methanol.

Measurement of motility and seizure-like behavior patterns

Planarian motility was measured by evaluating the swimming ability of the experimental animal in its natural state, using a petri dish (p-dish) filled with treatment solutions and divided into a grid of 0.5 cm intervals.



Fig. 1. The structure of microcystin-LR (A) and microcystin-LA (B).

Motility was evaluated by counting the number of grid lines that the planarians passed through [15]. Simultaneously, planarian seizure-like behaviors were also observed and the actions were classified into four categories: (A) snake-like, (B) screw-like, (C) c-like, and (D) head-bop [15]. Motility and seizure patterns were examined for 5 min, subsequent to a 5 min exposure to varying concentrations of MCLR or MCLA. Furthermore, planarians were also incubated in the absence/presence of 100 ng/mL MCLR or MCLA for 24, 48, and 96 hrs, after which they were transferred to clean water, left for 10 min to recover, with subsequent evaluation of motilities for 5 min.

Observation of eye formation in planarians

In order to examine the eye formation in planarians, the head region on the auricular grooves was cut, and rest of the body was incubated in water at 18° until emergence of eye extrusion. The process of eye formation was observed for 9 days, under a stereomicroscope (S8APO, Leica, Seoul, Korea).

Data analysis

Data analyses were processed using one-way analysis of variance (ANOVA) using SAS package 9.4 (SAS Institute, Cary, NC, USA) in a completely randomized design. Duncan's multiple range test was used to compare values of individual treatment when the *F*-value was significant (p<0.05).

Results

MCLR and MCLA attenuate the motility of planarians

Results were obtained after exposing planarians to vary-

ing concentrations of MCLR or MCLA for 5 min. Planarians were subsequently transferred to water, and motility was observed for 5 min. As presented in Fig. 2A, the number of motility decreased in planarians treated with MCLR or MCLA, as compared to controls, with no significant difference between groups. On the other hand, seizure-like behaviors were dose-dependently and significantly increased in planarians exposed to MCLR or MCLA (p<0.05, Fig. 2B). In particular, planarians treated with 10 or 100 ng/mL MCLA showed significantly higher incidence of screw type and head-bop type, as compared to other groups. Our results indicate that exposure to microcystins affects the motility and induces abnormal behaviors in planarians.

Exposure time of MCLR and MCLA affect to the behavior of planarians

In this study, planarians were incubated in water containing 100 ng/mL MCLR or MCLA for 0-96 hrs, with subsequent examination of motility and behavior (the control group was incubated in water containing methanol for 96 hrs). In Fig. 3A, the number of motility deceased in planarians exposed to MCLR or MCLA at each exposure time; especially, MCLA-treated planarians showed significantly decreased motility with prolonged exposure (p < 0.05). Moreover, the pattern of seizure-like behavior increased with increasing exposure to MCLR or MCLA, as compared to control group (Fig. 3B). The screw and head-bop characteristics were highly induced in planarians treated with MCLR for 96 hrs, whereas all seizure-like behaviors, including c-like, snake, screw and head-hop, were observed to be highly increased in planarians incubated with MCLA for 48 and 96 hrs (Fig. 3B, p < 0.05). Consequently, longer exposure to microcystin resulted in increased seizure behaviors and decrease mo-



Fig. 2. Effect of microcystins on the motility of planarians. Number of the motility (A) and seizure-like behavior (B) were counted in planarians after exposure to varying concentrations of microcystin-LR or -LA (control: methanol). Values are expressed as Mean \pm SEM. (^{a, b}) The different superscripts in each group of columns denote a significant difference at p<0.05.



Fig. 3. Motility patterns in planarians exposed to microcystin-LR or -LA. Number of the motility (A) and seizure-like behavior (B) were counted in planarians after exposing to different incubation times of microcystin-LR or -LA (control: methanol). Values are expressed as Mean \pm SEM. (^{a-c}) The different superscripts in each group of columns denote a significant difference at p < 0.05.

tility pattern in planarians, and showed a slightly more sensitive response to MCLA than MCLR.

Eye formation in the absence/presence of microcystin

In order to examine the regeneration pattern, the head region on the auricular grooves was decapitated, and rest of the body was incubated in water in the absence or presence of microcystin at 18°C, until eye extrusion was observed. As shown in Fig. 4, eye spot was observed simultaneously in all groups on day 4, and the eyes were completely formed on day 9, suggesting that microcystin does not affect eye formation, the most sensitive organ of the planarians.

Discussion

The environment is impacted most adversely with progressive revolutions in almost all fields in the world. Typically, it is the natural eco-system and organisms living in it that face maximum hardships due to the resulting pollution generated by humans. Natural water bodies are key centers of food chains and also for humans, since they provide water for the day to day activities [2, 16, 17].

Cyanobacteria, also called blue green algae, belong to the gram-negative bacterial group. They normally inhabit water reservoirs, and are engaged in photosynthesis. Pollution of water due to human activities results in increasing certain nutrient contents in the water, which ultimately leads to excessive growth, often defined as bacterial or algal blooming. Since cyanobacteria produce toxic compounds as their secondary metabolites, the increasing population of cyanobacteria results in higher toxic accu-

Incubation period
Day 1 Day 4 Day 9

 Jugged Horizon
 Day 1
 Day 4
 Day 9

 Jugged Horizon
 A
 A
 A

 Jugged Horizon
 A
 A
 A

 Jugged Horizon
 B
 B
 B
 B

 Jugged Horizon
 B
 B
 B
 B

 Jugged Horizon
 C
 C
 C
 C

Fig. 4. Eye formation in the absence/presence of microcystin. Amputated planarians were incubated in water in the absence (control; methanol, A-A") or presence of 100 ng/mL microcystin-LA (MCLA, B-B') or microcystin-LR (C-C") for 9 days. Eyes were observed from day 4 (black arrows, A', B' & C'), and were completely formed in planarians treated with or without microcystin at day 9 (observation magnification of stereo-microscope at \times 20).

mulation in the natural water resources. Consuming or living near such water bodies causes health problems to both humans and other living beings [1, 3, 4, 18]. Among the different toxic compounds, we selected microcystin to conduct the present study.

Microcystin is a substance that is continuously monitored due to concerns regarding its toxicity. In particular, controversy over the safety level of drinking water continues, and it has been designated and managed as a water quality inspection item in Korea since 2012 [19]. Microcystin can enter aquatic organisms and indirectly affect people who consume them. Moreover, exposure to low doses for prolonged periods of time leads to poisoning [20].

Planarian belongs to the group invertebrates; they have a flat body shape. Rearing them is quite easy due to their easy breeding and management requirements. They have a highly experimental value because of their well- developed central nervous system, similar to humans [13, 14]. Planarians are known to be the most sensitive to chemicals in their behavior, regeneration and brain structure, as compared to other alternative toxic animal models such as zebrafish, larvae and nematodes [21]. In addition, recent reports have suggested that planaria are useful in evaluating the toxicity of heavy metals such as copper and lead, and can be used to monitor water pollution [22].

Results of the present study reveal that motility and seizure patterns are negatively affected by both MCLR and MCLA. In some cases, the highest concentrations of microcystin showed reduction in the motility as well as increased seizure patterns in planarian (Fig. 2). Furthermore, both motility and seizure patterns are negatively affected with increasing incubation times with both MCLR and MCLA (Fig. 3). Normally, moderate or higher concentration of MCLR promotes reactive oxygen species (ROS), and subsequently results in oxidative DNA damage and transient DNA strand break [23]. The basic mechanism of this microcystin toxicity is probably the inhibition of protein phosphatases, followed by loss of cytoskeletal integrity and subsequent cytolysis or apoptosis (primarily of hepatocytes) [24, 25]. Microcystin toxicity has been reported to affect not only the liver but also the brain nerve. The neurotoxic effects by microcystin cause a variety of symptoms, namely, neuronal channels, signaling, oxidative stress, and cell skeletal disruption [26]. In this study, it was found that planarians treated with MCLR or MCLA caused more seizure-like behavior compared to motility pattern. Detailed mechanisms should be further studied in the future, but since planaria has a well-developed nervous system, it is thought that neurological seizures are more represented than motility. Previous studies provides clear evidence on the toxic biochemical reactions that occur in a living cell, which ultimately causes abnormalities in the living organism. Thus, abnormalities observed in motility, seizure and the regeneration process in the current study can possibly be attributed to alterations in the normal biochemical reactions due to the toxins.

Taken together, results of the present study indicate that there is a negative impact on the normal physical functioning of planarian, subsequent to microcystin exposure. This implies that the global increase in pollution poses a health risk to both humans and other living beings, as they have to live in an environment which is dependent on natural resources. Further studies are required to determine and express contrasting outcomes of this research area of interest.

Conflict of Interest

No potential conflict of interest relevant to this article was reported.

Acknowledgments

This research was supported by Sunchon National University Research Fund in 2020 (2020-0217). We thank Hee-Jung Lee and Changjung Kim, Jeonbuk National University, Korea.

ORCID

Young-Joo Yi, https://orcid.org/0000-0002-7167-5123 Yeon-Hwa Kim, https://orcid.org/0000-0002-4374-1963 A. A. Dilki Indrachapa Adikari, https://orcid.org/0000-0002-1021-3814 Seung-Tae Moon, https://orcid.org/0000-0003-1927-5250 Sang-Myeong Lee, https://orcid.org/0000-0002-3624-3392

Ethics Approval

This article does not require IRB/IACUC approval because there are no human and animal participants.

References

- Newcombe G, Chorus I, Falconer I, Lin TF. Cyanobacteria: impacts of climate change on occurrence, toxicity and water quality management. Water Res 2012;46:1347-1348.
- Briand JF, Jacquet S, Bernard C, Humbert JF. Health hazards for terrestrial vertebrates from toxic cyanobacteria in surface water ecosystems. Vet Res 2003;34:361-377.
- Tyagi MB, Thakur JK, Singh DP, Kumar A, Prasuna EG, Kumar A. Cyanobacterial toxins: the current status. J Microbiol Biotechnol 1999;9:9-21.
- Pitois S, Jackson MH, Wood BJB. Problems associated with the presence of cyanobacteria in recreational and drinking waters. Int J Environ Health Res 2000;10:203-218.
- Gassara F, Brar SK, Tyagi RD, Surampalli RY. Trends in biological degradation of cyanobacteria and toxins. In: Malik A, Grohmann E (eds.). Environmental protection strategies for sustainable development. Dordrecht, The Netherlands:

Springer; 2012. p. 261-294.

- Dietrich D, Hoeger S. Guidance values for microcystins in water and cyanobacterial supplement products (blue-green algal supplements): a reasonable or misguided approach? Toxicol Appl Pharmacol 2005;203:273-289.
- Pitois S, Jackson MH, Wood BJ. Sources of the eutrophication problems associated with toxic algae: an overview. J Environ Health 2001;64:25-32.
- Botes DP, Wessels PL, Kruger H, Runnegar MTC, Santikarn S, Smith RJ, Barna JCJ, Williams D H. Structural studies on cyanoginosins-LR, -YR, -YA, and -YM, peptide toxins from *Microcystis aeruginosa*. J Chem Soc Perkin Trans 1985;1:2747-2748.
- Botes DP, Tuinman AA, Wessels PL, Viljoen CC, Kruger H, Williams DH, Santikarn S, Richard J, Smith RJ, Hammond SJ. The structure of cyanoginocin-LA, a cyclic hepatopeptide toxin from the cyanobacterium *Microcystis aeruginosa*. J Chem Soc Perkin Trans 1984;1:2311-2318.
- de la Cruz AA, Antoniou MG, Hiskia A, Pelaez M, Song W, O'Shea KE, He X, Dionysiou DD. Can we effectively degrade microcystins? - implications on human health. Anticancer Agents Med Chem 2011;11:19-37.
- Bell SG, Codd GA. Cyanobacterial toxins and human health. Rev Med Microbiol 1994;5:256-264.
- Van Apeldoorn ME, Van Egmond HP, Speijers GJ, Bakker GJ. Toxins of cyanobacteria. Mol Nutr Food Res 2007;51:7-60.
- Raffa RB. Planaria: a model for drug action and abuse. Boca Raton: CRC Press; 2008.
- Ross KG, Currie KW, Pearson BJ, Zayas RM. Nervous system development and regeneration in freshwater planarians. Wiley Interdiscip Rev Dev Biol 2017;6:e266.
- Kim YJ, So J, Yi YJ, Lee SM. Effect of glycyrrhizin on nicotine-induced behavioral disturbance of planarian, *Dugesia japonica*. J Biomed Transl Res 2019;20:21-29.
- Palma P, Alvarenga P, Palma V, Matos C, Fernandes RM, Soares A, Barbosa IR. Evaluation of surface water quality using an ecotoxicological approach: a case study of the Alqueva Reservoir (Portugal). Environ Sci Pollut Res 2010;

17:703-716.

- 17. da Silva CA, Oba ET, Ramsdorf WA, Magalhães VF, Cestari MM, Ribeiro CAO, de Assis HCS. First report about saxitoxins in freshwater fish *Hoplias malabaricus* through trophic exposure. Toxicon 2011;57:141-147.
- Chorus I, Bartram J. Toxic cyanobacteria in water: a guide to their public health consequences, monitoring and management. Boca Rataon: CRC Press; 1999.
- Kim DS, Kim SH, Kim HJ, Baek OJ, Kang YW, Lee JG, Cho MJ, Shin CS. Studies on the validation of analytical method for microcystins in freshwater fish. Ministry of Food and Drug Safety, Korea. 2015.
- Ibelings BW, Chorus I. Accumulation of cyanobacterial toxins in freshwater "seafood" and its consequences for public health: a review. Environ Pollut 2007;150:177-192.
- Hagstrom D, Cochet-Escartin O, Zhang S., Khuu C, Collins EMS. Freshwater planarians as an alternative animal model for neurotoxicology. Toxicol Sci 2015;147:270-285.
- Prá D, Lau AH, Knakievicz T, Carneiro FR, Erdtmann B. Environmental genotoxicity assessment of an urban stream using freshwater planarians. Mutat Res Genet Toxicol Environ Mutagen 2005;585:79-85.
- Žegura B, Sedmak B, Filipič M. Microcystin-LR induces oxidative DNA damage in human hepatoma cell line HepG2. Toxicon 2003;41:41-48.
- Wickstrom ML, Khan SA, Haschek WM, Wyman JF, Eriksson JE, Schaeffer DJ, Beasley VR. Alterations in microtubules, intermediate filaments, and microfilaments induced by microcystin-LR in cultured cells. Toxicol Pathol 1995;23:326-337.
- 25. Fischer WJ, Hitzfeld BC, Tencalla F, Eriksson JE, Mikhailov A, Dietrich DR. Microcystin-LR toxicodynamics, induced pathology, and immunohistochemical localization in livers of blue-green algae exposed rainbow trout (*Oncorhynchus mykiss*). Toxicol Sci 2000;54:365-373.
- Hu Y, Chen J, Fan H, Xie P, He J. A review of neurotoxicity of microcystins. Environ Sci Pollut Res 2016;23:7211-7219.