Original Article

Therapeutic effects of combination of *Galla rhois* extract and Sodium chlorate on Mice infected with *Brucella abortus*

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This study investigated the therapeutic effects of Galla rhois (GR) ethanol extract (GRE), sodium chlorate (SC), and a combination of GRE and SC on mice infected with Brucella abortus (B. abortus). Mice were infected intraperitoneally with B. abortus and then treated with GRE, SC, and a combination GRE and SC in drinking water for 14 days. Then, serum antibodies were used in a tube agglutination test (TAT), after which the weight and CFUs from each spleen were measured. In addition, histopathological changes in each liver were examined at 14 days postinfection. At 14 days post-infection, negative reactions of serum antibodies in PC (positive control), SCT (SC 1.6 g/L drinking water), GRT (GRE 200 mg/L drinking water), and GST (GRE 200 mg + SC 1.6 g/L drinking water) were 0, 40, 60, and 80%, respectively. The average spleen weight was not significantly different between the groups. At 14 days post-infection, bacterial numbers in all treated groups were significantly lower compared to to that of the PC (GRT and SCT, P<0.05; GST, P<0.001). In terms of histopathological changes in the livers, there were numerous multifocal microgranulomas in the PC, whereas this number successively decreased in the SCT, GRT, and GST groups. Conclusively, a combination of GRE and SC exhibits therapeutic effects on mice infected with B. abortus. These results suggest the potential efficacy of a mixture of GRE and SC in the treatment of brucellosis.

Key words: *Brucella abortus, Galla rhois,* sodium chlorate, combination, mice

Introduction

Brucellosis is a major zoonosis in domestic animals and

causes economic losses for the livestock industry as well as public health problems for humans [27]. The disease is caused by *Brucella* spp., which are small, Gram-negative, non-motile and polymorphic rods. *Brucella* spp. are facultative intracellular parasites that cause abortion and infertility in various mammals and the well-known undulant fever in humans [9]. The bacteria penetrate the mucosa of the nasal, oral, or pharyngeal cavities and are phagocytized by host macrophages, where survival and replication occurs [12].

Currently, combinations of two or three antibiotics are used to treat brucellosis due to the low efficacy of monotherapy. Several clinical studies analyzed the efficiency of different antibiotic regimens, and identified problems such as financial considerations in developing countries, therapeutic failures, relapses and the emergence of drug resistance [4, 32].

In general, tetracycline/aminoglycoside combinations are the most common antibiotics used in the treatment of brucellosis [31]. However, due to high rates of treatment failure or relapses due to emerging drug resistance, antibiotic treatment of brucellosis is still problematic [24]. Thus, alternative therapies to treat brucellosis are needed.

Conventional herbal medicines have long been used as remedies against infectious diseases in Asian countries. *Galla rhois* (GR) has long been used in traditional Asian medicine to treat diarrhea, persistent coughing and spontaneous perspiration due to its antidiarrhetic, astringent and hemostatic properties [10, 13, 22, 23]. GR is a harmless natural material that contains a number of tanninderived components, including methyl gallate and gallic acid [15]. Notably, the gallotannins are a class of hydro-

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lysable tannin polymers formed from gallic acid, which seems to have anti-bacterial, anti-fungal, and anti-viral properties [3, 14]. In a previous study [22], GR ethanol extract exhibited antibacterial and protective activities against *Brucella abortus* (*B. abortus*) *in vitro* and *in vivo*.

Sodium chlorate (SC) is used as an oxidizing agent and for making chlorine dioxide used in water disinfection. Chlorate is found as a stable by-product in drinking water that has been disinfected with chlorine dioxide [25]. Previous studies have been carried out on the prevention and treatment of *Enterobacteriaceae* infections in animals using SC [6, 7]. *B. abortus* like *Salmonella* species has a respiratory nitrate reductase enzyme, which coincidentally catalyses the intracellular reduction of chlorate into chlorite, a cytotoxic product that kills the bacterium in tissue cells [8, 11]. As most of the normal anaerobic gut bacteria lack respiratory nitrate reductase activity, chlorate selectively targets bacteria expressing respiratory nitrate reductase activity but not beneficial anaerobes lacking that enzyme [6, 7].

Although previous studies [6-8, 22] investigated the antibacterial effects of each of GR and SC, few studies have been carried out to investigate the effect of a combination of GR and SC on mice infected with *B. abortus*. The present study evaluated the therapeutic potential of a combination of GR ethanol extract (GRE) and SC on murine brucellosis.

Materials and Methods

GRE and SC solution preparation

GR powder was obtained from GS Bio (Jeonju, Korea), isolated from plant material and analyzed as described previously [2, 22]. Briefly, 1 kg of plant material was dried in an oven at 60°C for 3 days and extracted with ethanol twice at room temperature. The remaining residue was removed by filtration (Whatman no. 2, Sigma-Aldrich Korea, Yonin), and the filtrate was concentrated using a vacuum rotary evaporator (Iwai Co., Japan), followed by freezing of the dried powder. This crude, extracted powder was used in the present study.

SC was purchased from Sigma-Aldrich Korea (Yonin, Korea). To make a 30 mM SC stock solution, 3.19 g of SC was dissolved in distilled water up to a final volume of 1L. After the SC stock solution was suitably diluted with distilled water, the diluents were used in this study.

Animals

Six-week-old specific pathogen-free (SPF) female ICR mice were obtained from Samtaco Co. (Osan, Korea). All animals were kept at the inspecting facility of Gyeong-sang National University (Chinju, Korea) for 1 week to allow acclimation before experimentation. Thereafter, they were kept in an isolated SPF barrier room with

regulated temperature $(23 \pm 1^{\circ}C)$, humidity $(50 \pm 5\%)$ and light/dark cycle (12/12 hr). The animals were fed a sterilized (2 M rad radiation) pellet diet (Purina, Seoul, Korea) and sterilized water *ad libitum*. All studies were performed in accordance with the Guide for Animal Experimentation of Gyeongsang National University and approved by the Institutional Animal Care and Use Committee of Gyeongsang National University (Approval No. GNU-2013-12-10). All efforts were made to minimize pain or discomfort experienced by the used animals.

Bacterial strains and culture conditions

B. abortus strains were derived from 544 (ATCC 23448), a smooth, virulent *B. abortus* biovar 1 strain. *B. abortus* strains were maintained as frozen glycerol stocks and were cultured in Brucella broth (Becton Dickinson, Sparks, MD.) or Brucella broth containing 1.5% agar without antibiotics for 3 days at 37°C. Bacteria were grown at 37°C with vigorous shaking until they reached the stationary phase, and bacterial growth rates were measured using a spectrophotometer (Beckman Coulter Korea, Seoul) at a wavelength of 600 nm.

Experimental design

After a 1-week adaptation period, all mice were injected intraperitoneally with 2×10^4 CFUs of *B. abortus*. Similar to previous studies [6-8, 22], forty ICR mice infected with *B. abortus* were randomly divided into four groups: PC (Positive control), GRT (GRE 200 mg/L drinking water), SCT (sodium chlorate 1.6 g/L drinking water), and GST (GRE 200 mg + SC 1.6 g/L drinking water). All mice were fed drinking water treated with each drug *ad libitum* for 14 days.

Titration of antibodies to B. abortus

On day 7 and 14 after drug treatment, five mice from each group were sacrificed, and their bloods were collected, and centrifuged at $1,000 \times g$ for 30 min to separate the serum. According to previous protocols for the tube agglutination test (TAT) [26, 29], the diagnostic antigen for *B. abortus*, supplied by the Animal and Plant Quarantine Agency (Anyang, Korea), was diluted at 1:100 in phenol saline before use. Thereafter, 0.08, 0.04, 0.02, 0.01, and 0.005 mL of serum samples, inactivated at 56°C for 30 min, were placed in different tubes and mixed with 2 mL of the diluted antigen. The results were read after incubation at 37°C for 48 hr. The criteria for positive, suspected positive and negative reactions were greater than 100, 50, and less than 25 serum dilution, respectively [20, 33].

Bacteriological examination of the spleen

On day 7 and 14 post-infection, five mice from each group were sacrificed, and their spleens were removed and weighed. In addition, each spleen was homogenized

in PBS. Continually, the homogenates were serially diluted with PBS and plated on Brucella agar. After the plates were incubated at 37°C for 3 days, the number of CFUs in each spleen was counted and represented as CFU/g spleen.

Histopathological examination of the liver

At day 14 post-infection, five mice from each group were sacrificed, and their livers were removed. The livers were fixed in 10% neutral-buffered formalin for at least 24 hr. Tissues were dehydrated in graded alcohols, cleared with xylene, and infiltrated and embedded in paraffin. Tissues embedded in paraffin were cut to a thickness of $4~6 \mu m$ and mounted on glass slides. The sections were stained with hematoxylin and eosin (H&E) and examined for histopathological changes under a light microscope (Olympus, Tokyo, Japan).

Statistical analysis

The results obtained were expressed as the means \pm S.D. for the replicate experiments. Student's *t*-test was used to make a statistical comparison between the groups. Results with *P*<0.05 were considered statistically significant.

Results

Titration of serum antibodies

To identify the inhibitory effects of GRE, SC and a mixture of GRE and SC, indicated by a decrease in the ability of *B. abortus* to promote infection within the host, mice infected with *B. abortus* were treated with GRE, SC and a mixture of GRE and SC, and serum antibodies of *B. abortus* were titered at day 7 and 14 post-infection.

The reciprocal antibody titers from TAT in sera for the ICR mice infected with *B. abortus* are presented in Table 1. At 7 days post-infection, the antibody titers in PC and SCT ranged from 1:50 to 1:100 \leq and those in GRT and GST ranged from \leq 1:25 to 1:100 \leq . At 14 days post-infection, the antibody titers ranged from 1:200 to 1:400 in PC, from \leq 1:25 to 1:100 \leq in SCT and from \leq 1:25 to 1:50 in GRT and GST. In GRT and GST at 7 and 14 days post-infection, the reciprocal antibody titers showed a tendency to decrease compared to NC.

Spleen weight

The average spleen weight in PC, GRT, SCT and GST was 0.25 ± 0.05 g, 0.18 ± 0.04 g, 0.21 ± 0.04 g and 0.16 ± 0.03 g, respectively. In GRT, SCT and GST at 14 days post-infection, the average spleen weight decreased compared to those in PC (Table 2). However, there was no significant difference between groups.

Number of B. abortus CFUs per spleen

Fig. 1 shows the number of bacteria recovered from the spleens in each group at 7 and 14 days post-infection. At 7 days post-infection, there was a significant difference between PC and GST (P<0.05). At 14 days post-infection, bacterial numbers in all treated groups were significantly decreased compared to PC (GRT and SCT, P<0.05; GST, P<0.001). These findings indicate that GRE, SC and a combination of GRE and SC have a therapeutic effect

Table 1. Results of the antibody titers from tube agglutination tests for the sera of mice infected with B. abortus

Group	After treatment	Negative ≤1:25	Suspected positive 1:50	Positive		
				1:100≤	1:200	1:400
РС	Day 7		2	3		
	Day 14				1	4
SCT	Day 7		2	3		
	Day 14	2	2	1		
GRT	Day 7	1	2	2		
	Day 14	3	2			
GST	Day 7	1	2	2		
	Day 14	4	1			

Table 2. Average sp	bleen weight for each	h group on Day 14	4 post-infection
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T4	Group				
Item	РС	SCT	GRT	GST	
Weight (g)	0.25 ± 0.05	0.21 ± 0.04	0.18 ± 0.04	0.16 ± 0.03	

on murine brucellosis, and the mixture of GRE and SC shows the most curative effect.

Histopathological examination of the liver

To examine the pathologic changes from the formation of microgranulomas containing inflammatory cells, slides were prepared from the livers of mice infected with *B. abortus*, and observed under a light microscope. In the mice livers, there were numerous multifocal microgranulomas in PC, and their number decreased successively in SCT, GRT and GST (Fig. 1).

Discussion

Brucellosis is one of the world's most widespread zoonoses and is of major economic importance in most countries around the world [5]. The disease has been efficiently controlled in agricultural animals through the use of vaccination, surveillance and confined programs [22]. However, there is currently no safe or efficient vaccine that can be recommended to control brucellosis [19]. In addition, treatment of the disease with antibiotics remains controversial and requires prolonged therapy with at least two agents [28]. Moreover, Brucella organisms escape the host's defense system, as the bacteria invade, resist killing, replicate and appear to be well-adapted to endure the multiple environmental conditions in an intracellular niche inside the host [21]. Therefore, conventional antibiotic regimens are not suitable for the treatment of Brucella infection due to its escape from the host's defense system and the rapid emergence of antibiotic resistance [1].

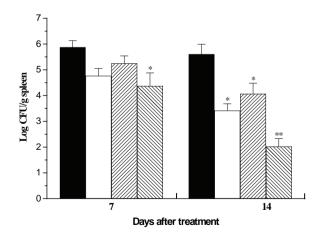


Fig. 1. The number of *B. abortus* isolated from the spleens of infected mice. Control (**u**): untreated mice; GRT (\Box): mice treated with GRE at a concentration of 200 mg/L drinking water; SCT (\boxtimes): mice treated with SC at a dose of 1.6 g/L drinking water; and GST (\boxtimes), mice treated with a combination of 200 mg/L and 1.6 g/L drinking water. *, **Significant difference at *P*<0.05 and *P*<0.001 levels, respectively, compared with the control group.

In previous studies, research on alternatives to conventional antibiotics to treat brucellosis using natural products has not been widely reported. This study was carried out to determine the potential of GRE, SC and a combination of GRE and SC as alternatives to antibiotics for brucellosis treatment.

In this study, the negative results of serum antibody titers in GRT and GST were 60% (3/5) and 80% (4/5) after 14-day treatment, respectively, while the positive result of those in NC was 100% at 14 days post-infection. In the antibiotic treatment of canine brucellosis, all infected dogs given 5 mg/kg of enrofloxacin orally every 12 hr for 30 days were reported negative on the serological rapid slide agglutination test [34]. In addition, the results of antibiotic treatment for 6 weeks in patients with brucellosis were reported positive in 20% of patients by serum plate agglutination test (SPA), while the SPA results were positive in 80% of patients before antibiotic treatment [16]. Considering the infected subjects and the treatment period, GRE and a combination of GRE and SC in this study were more therapeutic than the antibiotics in the above studies.

The liver is an important site for colonization and replication of *Brucella* in mice. Usually, mice infected with virulent strains of *Brucella* spp. have mild to moderate hepatitis, which is characterized by neutrophilic infiltrate in the early stages of infection, followed by histiocytic infiltrate with epithelioid cells and microgranulomas in the chronic stages of infection [18, 30]. In this study, mi-

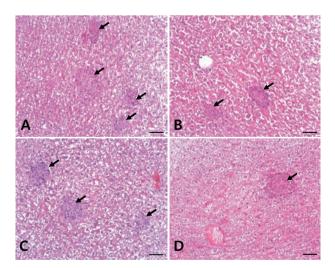


Fig. 2. Representative photographs of liver sections. (A) A control mouse showing numerous microgranulomas (arrows) containing inflammatory cells. (B) A GRE-treated mouse showing a moderate decrease in microgranulomas compared to the untreated-control mouse. (C) A SC-treated mouse showing a mild decrease in microgranulomas. (D) A combination of GRE and SC treated mouse showing a severe decrease in microgranulomas. H&E stain. Scale bar: 50 µm.

crogranulomas were found in the liver of mice infected with *B. abortus* in NC at 14 days post-infection. In addition, microgranulomas gradually decreased in SCT, GRT and GST, which indicate that SC, GRE and a combination of GRE and SC have a therapeutic effect on murine brucellosis.

To examine the effects of the treatment of SC, GRE and a mixture of GRE and SC, mice were infected with B. abortus. In SCT, GRT and GST, the average weight of spleens collected from infected mice showed no significant difference compared to those in PC. However, the number of B. abortus in the spleens in GRT (P<0.05) and GST (P<0.001) was significantly decreased compared to that in PC, and the bacterial loads in the spleens of mice in GRT and GST after 14 days post-treatment were more than 150- and 3,000-fold lower than those of PC, respectively (Fig. 1). These results indicated that a combination of GRE and SC has a potential therapeutic effect on murine brucellosis. In a previous study on the treatment of murine brucellosis with antibiotics [17], erythromycin was orally administered to infected mice at a concentration of 200 mg/kg/day for 15 days post-infection, and the number of B. abortus recovered from the spleens of the treated mice was approximately 16-fold lower than that in the untreated mice. Taking into account the bacteria decrease in the spleens of mice, GRE and the combination of GRE and SC in this study have a greater potential therapeutic effect than erythromycin.

Conclusively, this study emphasizes the idea that the combination of GRE and SC is effective in the treatment of brucellosis, and possibly other diseases caused by intracellular pathogenic bacteria, and is an alternative to conventional therapy regimes.

In the future, it will be necessary to determine the clinical usefulness of a mixture of GRE and SC in the treatment of brucellosis.

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References

- Adesiyun AA, Baird K, Stewart-Johnson A. Antimicrobial resistance, phenotypic characteristics and phage types of *B. abortus* strains isolated from cattle and water buffalo (*Bubalus bubalis*) in Trinidad. Vet Arhiv 2011;81:391-404.
- Ahn YJ, Lee CO, Kweon JH, Ahn JW, Park JH. Growthinhibitory effects of *Galla Rhois*-derived tannins on intestinal bacteria. J Appl Microbiol 1998;84:439-443.

- Ahn YJ, Lee HS, Oh HS, Kim HT, Lee YH. Antifungal activity and mode of action of *Galla Rhois*-derived phenolics against phytopathogenic fungi. Pestic Biochem Phys 2005;81:105-112.
- Al Dahouk S, Nockler K. Implications of laboratory diagnosis on brucellosis therapy. Expert Rev Anti-infect Ther 2011;9:833-845.
- Alemayehu A. Review on emerging and re-emerging bacterial zoonotic diseases. Am-Euras J Sci Res 2012;7:176-186.
- Anderson RC, Buckley SA, Kubena LF, Stanker LH, Harvey RB, Nisbet DJ. Bactericidal effect of sodium chlorate on *Escherichia coli* O157:H7 and *Salmonella* Typhimurium DT104 in rumen contents *in vitro*. J Food Prot 2000;63:1038-1042.
- Anderson RC, Harvey RB, Byrd JA, Callaway TR, Genovese KJ, Edrington TS, Jung YS, McReynolds JL, Nisbet DJ. Novel preharvest strategies involving the use of experimental chlorate preparations and nitro-based compounds to prevent colonization of foodproducing animals by foodborne pathogens. Poult Sci 2005;84:649-654.
- Anderson RC, Hume ME, Genovese KJ, Callaway TR, Jung YS, Edrington TS, Poole TL, Harvey RB, Bischoff KM, Nisbet DJ. Effect of drinking water administration of experimental chlorate ion reparations on *Salmonella* enteric serovar Typhimurium colonization in weaned and finished pigs. Vet Res Commun 2004;28:179-189.
- Arenas GN, Staskevich AS, Aballay A, Mayorga LS. Intracellular trafficking of *Brucella abortus* in J774 macrophages. Infect Immun 2000;68:4255-4263.
- Aziz NH, Farag SE, Mousa LA, Abo-Zaid MA. Comparative antibacterial and antifungal effects of some phenolic compounds. Microbios 1998;93:43-54.
- 11. Callaway TR, Anderson RC, Genovese KJ, Poole TL, Anderson TJ, Byrd JA, Kubena LF, Nisbet DJ. Sodium chlorate supplementation reduces *E. coli* O157:H7 populations in cattle. J Anim Sci 2002;80:1683-1689.
- Cardoso PG, Macedo GC, Azevedo V, Oliveira SC. Brucella spp noncanonical LPS: structure, biosynthesis, and interaction with host immune system. Microb Cell Fact 2006;5:13.
- Cha CN, Yu EA, Park EK, Choi H, Kim S, Lee HJ. Antibacterial effects of *Galla Rhois* extract against *Streptococcus suis* infection in mice. J Fd Hyg Safety 2013;28:95-98.
- Chen JC, Ho TY, Chang YS, Wu SL, Hsiang CY. Antidiarrheal effect of *Galla Chinensis* on the *Escherichia coli* heat-labile enterotoxin and ganglioside interaction. J Ethnopharmacol 2006;103:385-391.
- Djakpo O, Yao W. *Rhus chinensis* and *Galla Chinensis*folklore to modern evidence: review. Phytother Res 2010;24:1739-1747.

- Elfaki MG, Al-Hokai AA, Nakeeb SM, Al-Rabiah FA. Evaluation of culture, tube agglutination, and PCR methods for the diagnosis of brucellosis in humans. Med Sci Monit 2005;11:69-74.
- Felek S, Demirdag K, Kalkan A, Akbulut A. Therapeutic effects of rifampin and erythromycin in experimental murine brucellosis. Clin Microbiol Infec 2000;6:111-114.
- Grilló MJ, Blasco JM, Gorvel JP, Moriyón I, Moreno E. What have we learned from brucellosis in the mouse model?. Vet Res 2012;43:29.
- Harms JS, Durward MA, Magnani DM, Splitter GA. Evaluation of recombinant invasive, non-pathogenic *Escherichia coli* as a vaccine vector against the intracellular pathogen, *Brucella*. J Immune Based Ther Vaccines 2009;7:1.
- Kim CH, Bang SY, Jeon JH, Bhak JS, Lee MK, Shin JS. A comparison of agglutination test and enzyme-linked immunosorbent assay for the bovine brucellosis. Korean J Vet Serv 2008;31:315-329.
- Ko J, Splitter GA. Molecular host-pathogen interaction in brucellosis: current understanding and future approaches to vaccine development for mice and humans. Clin Microbiol Rev 2003;16:65-78.
- Lee JJ, Bae JH, Kim DH, Lim JJ, Kim DG, Lee HJ, Min W, Rhee MH, Chang HH, Park H, Kim S. Intracellular replication inhibitory effects of *Galla Rhois* ethanol extract for *Brucella abortus* infection. J Ethnopharmacol 2011;138:602-609.
- Lee JJ, Kim DH, Lim JJ, Kim DG, Min W, Kim GS, Lee HJ, Rhee MH, Park H, Kim SC, Chang HH, Kim S. Anticoccidial effect of supplemental dietary *Galla Rhois* against infection with *Eimeria tenella* in chickens. Avian Pathol 2012;41:403-407.
- 24. Motamedi H, Darabpour E, Gholipour M, Seyyed-Nejad SM. *In vitro* assay for the anti-brucella activity of

medicinal plants against tetracycline-resistant *Brucella melitensis*. J Zhejiang Univ Sci B 2010;11:506-511.

- Cha CN, Jung WC, Choi HC, Lee YE, Yoo CY, Kim S, Lee HJ. Effects of short-term sodium chlorate exposure on pigs. Acta Vet Hung 2012;60:93-101.
- Stemshorn BW, Forbes LB, Eaglesome MD, Nielsen KH, Robertson FJ, Samagh BS. A comparison of standard serological tests for the diagnosis of bovine brucellosis in Canada. Can J Comp Med 1985;49:391-394.
- Prior S, Gander B, Irache JM, Gamazo C. Gentamicinloaded microspheres for treatment of experimental *Brucella abortus* infection in mice. J Antimicrob Chemother 2005;55:1032-1036.
- Prior S, Gander B, Lecároz C, Irache JM, Gamazo C. Gentamicin-loaded microspheres for reducing the intracellular *Brucella abortus* load in infected monocytes. J Antimicrob Chemother 2004;53:981-988.
- Rahman MS. Plate and tube agglutination tests for diagnosis of *Brucella abortus* biotype 1 infection in Spraque-Dawley rats. Bangl J Vet Med 2004;2:63-67.
- Silva TM, Costa EA, Paixão TA, Tsolis RM, Santos RL. Laboratory animal models for brucellosis research. J Biomed Biotechnol 2011:2011;518323.
- Skalsky K, Yahav D, Bishara J, Pitlik S, Leibovici L, Paul M. Treatment of human brucellosis: systematic review and meta-analysis of randomised controlled trials. Br Med J 2008;336:701-704.
- 32. Solera J. Update on brucellosis: therapeutic challenges. Int J Antimicrob Agents 2010;36:S18-S20.
- 33. Sung SR, Kim JY, Her M, Lee K, Gu JH, Kang SI, Lee HK, Kim SM, Jung SC. Evaluation on diagnostic efficiency of the standard tube agglutination test for bovine brucellosis. Korean J Vet Serv 2012;35:269-273.
- Wanke MM, Delpino MV, Baldi PC. Use of enrofloxacin in the treatment of canine brucellosis in a dog kennel (clinical trial). Theriogenology 2006;66:1573-1578.